

Original article

## *Paracoccidioides restrepiensis* has undergone a severe population bottleneck

### *Paracoccidioides restrepiensis* ha experimentado un grave cuello de botella

✉ Juan G. McEwen<sup>1,2,\*</sup>, ✉ Oscar M. Gómez<sup>1,2</sup>, ✉ Daniel R. Matute<sup>3</sup>

<sup>1</sup> Corporación para Investigaciones Biológicas (CIB) Medellín, Colombia

<sup>2</sup> Facultad de Medicina, Universidad de Antioquia, Medellín, Colombia

<sup>3</sup> Biology Department, University of North Carolina, Chapel Hill, North Carolina, USA

Inaugural article of Juan Guillermo McEwen as corresponding member of the Colombian Academy of Exact, Physical and Natural Sciences

#### Abstract

Genus *Paracoccidioides* now encompasses five species, *P. lutzii*, *P. brasiliensis sensu stricto*, *P. americana*, *P. venezuelensis*, and *P. restrepiensis*. All the paracoccidioidomycosis (PCM) cases reported in Colombia have been classified as *P. restrepiensis*, which was named in honor of Angela Restrepo, a leading researcher in the field of the biology of the fungus. Previous assessments of genetic diversity have suggested that *Paracoccidioides* species have differences in their level of polymorphism. To infer changes in effective population size, we used the Pairwise sequentially Markovian coalescent (PSMC) by generating two pseudodiploids for *P. restrepiensis* and one for *P. brasiliensis sensu stricto*. We found that *P. restrepiensis* diverged from its sister species recently, the divergence time from *P. venezuelensis* being 125,000 ( $\pm$  42,000) years. The analyses using PSMC show a systematic reduction in the effective population size of *P. restrepiensis* with a rapid decrease in genetic variability compared to *P. brasiliensis sensu stricto*, which indicates that *P. restrepiensis* has undergone a systematic population bottleneck. None of the other two species show the dramatic effective population size reduction observed in *P. restrepiensis*. These comparisons suggest that the trajectory of *P. restrepiensis* is somehow different from the other *Paracoccidioides* species and poses the question regarding the biogeographic events that have led to such a dramatic population pattern.

**Keywords:** *Paracoccidioides* spp.; *Paracoccidioides restrepiensis*; Paracoccidioidomycosis; Effective population size.

#### Resumen

El género *Paracoccidioides* hoy abarca cinco especies: *P. lutzii*, *P. brasiliensis sensu stricto*, *P. americana*, *P. venezuelensis* y *P. restrepiensis*. Todos los casos de paracoccidioidomycosis reportados en Colombia han sido causados por *P. restrepiensis*, denominado así en honor de Ángela Restrepo, líder en el estudio de este hongo. Las evaluaciones previas de la diversidad genética han sugerido que las especies de *Paracoccidioides* difieren en su polimorfismo. Para inferir cambios en el tamaño efectivo de la población, se utilizó el método del coalescente markoviano secuencial por pares (*pairwise sequentially Markovian coalescent*, PSMC) mediante la generación de dos pseudodiploides para *P. restrepiensis* y uno para *P. brasiliensis sensu stricto*. Se encontró que *P. restrepiensis* divergió de sus especies hermanas recientemente, con un tiempo de divergencia en relación con *P. venezuelensis* de 125.000 ( $\pm$  42.000) años. Los análisis que utilizan el PSMC muestran una reducción sistemática en el tamaño efectivo de la población de *P. restrepiensis*, con una rápida disminución de la variabilidad genética en comparación con *P. brasiliensis sensu stricto*, lo que indica que *P. restrepiensis* se ha visto expuesto a un cuello de botella poblacional sistemático. Ninguna de las otras dos especies muestra la drástica reducción efectiva del tamaño de

**Citation:** McEwen JG, Gómez OM, Matute DR. *Paracoccidioides restrepiensis* has undergone a severe population bottleneck. Revista de la Academia Colombiana de Ciencias Exactas, Físicas y Naturales. 46(181):866-876, octubre-diciembre de 2022. doi: <https://doi.org/10.18257/raccefyn.1768>

**Editor:** Elizabeth Castañeda

**\*Corresponding autor:**

Juan G McEwen; [mcewen@une.net.co](mailto:mcewen@une.net.co)

**Received:** September 13, 2022

**Accepted:** November 3, 2022

**Published on line:** November 24, 2022



This is an open access article distributed under the terms of the Creative Commons Attribution License.

la población observada en *P. restrepiensis*. Estas comparaciones sugieren que la trayectoria de *P. restrepiensis* es, de alguna manera, diferente de las otras especies de *Paracoccidioides* y llevan a preguntarse cuáles han sido los eventos biogeográficos que han producido cambios tan dramáticos en el patrón poblacional.

**Palabras claves:** *Paracoccidioides* spp., *Paracoccidioides restrepiensis*; *Paracoccidioidomycosis*; Tamaño de población efectivo.

## Introduction

*Paracoccidioides* spp. is a genus of temperature-dependent dimorphic fungi that causes paracoccidioidomycosis (PCM), a neglected tropical systemic mycosis. Brazil reports 80 % of the cases with an estimated incidence of 3 to 4 new cases/million and up to 8 to 10 cases/million in highly endemic areas (Bellissimo-Rodrigues *et al.*, 2011; Coutinho *et al.*, 2015). Since 1908, over 15,000 cases have been reported. The mycosis exhibits two main clinical presentations: the subclinical, or asymptomatic infection, and the clinically manifested disease. The latter is usually chronic and involves the primary target, i.e., the lungs, as well as the mucosa and different organs such as the skin, the adrenal glands, and lymph nodes, among others. PCM affects adult males at least ten times more than women, with a male-female ratio of approximately 11:1 (Restrepo-Moreno *et al.*, 2020). The disease has long latency periods, up to 30 years in some cases, as evidenced by cases reported outside the endemic area (Restrepo, 2000).

The first two cases of PCM were reported in 1908 by Adolpho Lutz (1855-1940) (Lutz, 1908, 2004). In 1912, Alfonso Splendore (1871-1953), a colleague of Dr. Lutz, reported four new cases of the disease and placed the fungus in the genus *Zymonema*, as *Zymonema brasiliensis* (Splendore, 1912). In 1930, Floriano Paulo de Almeida (1898-1977) published a comparative study of *Coccidioides immitis* and this new agent demonstrating that PCM and coccidioidomycosis were two different diseases and classifying the agent as *Paracoccidioides brasiliensis* (Almeida, 1930). A consensus on the name and classification was finally achieved in 1971 at the PCM symposium in Medellín, Colombia (PAHO, 1971).

Initially, the disease was named Brazilian blastomycosis, then South American blastomycosis, Lutz disease, and Lutz-Splendore-Almeida disease, among other names. Today, PCM is the universally accepted denomination (Franco *et al.*, 1994).

Since its discovery, *Paracoccidioides* has been the focus of intense research, but despite the exquisite characterization of the immune response and the biochemical characteristics of *Paracoccidioides*, the ecology and evolutionary biology of the fungus remained largely unexplored until the advent of molecular biology. The use of gene genealogies and later the use of wide phylogenetic reconstructions of the genome revealed the existence of species boundaries within what was previously thought to be a single species. The genus *Paracoccidioides* now encompasses *P. lutzii*, *P. brasiliensis sensu stricto*, *P. americana*, *P. venezuelensis*, and *P. restrepiensis*. The five species show phenotypic differences in their morphology (Turissini *et al.*, 2017), pathogenesis (Scorzoni *et al.*, 2018), and the immune response elicited in the host (Assolini *et al.*, 2021; Siqueira *et al.*, 2016), although there has been no systematic assessment of the phenotypic differences among all species.

Notably, the species have distinct yet overlapping geographic ranges suggesting that opportunities for hybridization and gene exchange do exist (Matute *et al.*, 2006) The most prevalent species in Colombia was named *P. restrepiensis* in honor of Angela Restrepo, a trailblazer in the study of the biology of the fungus (Turissini *et al.*, 2017). All the PCM cases reported in Colombia and subjected to molecular characterization have been classified as *P. restrepiensis* suggesting that this species is particularly important in Northern South America (Matute *et al.*, 2006).

The extent of genetic variation in *Paracoccidioides* has been studied since the advent of molecular biology techniques in the 1990s. Initial surveys with RAPDs and RFLPs revealed variations but didn't ascribe them to species boundaries (Morais *et al.*, 2000;

Nino-Vega *et al.*, 2000; Soares *et al.*, 1995). The first formal genetic efforts to characterize the extent of genetic variation in *P. brasiliensis sensu lato* populations used Sanger sequencing to detect genetic polymorphism in short amplicons and revealed the existence of four genetically isolated clades that satisfied the definition of phylogenetic species. Later, these approaches revealed the existence of five different genetically isolated clades that satisfied the definition of phylogenetic species (Matute *et al.*, 2006; Teixeira *et al.*, 2009; Turissini *et al.*, 2017).

Genome sequencing and population genomics allowed for more formal speciation tests (Matute & Sepulveda, 2019). However, the study of short sequences is inherently limited because it can only reveal the history of individual genes while the incorporation of genome-wide polymorphism has provided the opportunity of collating multiple gene genealogies and extracting the most likely species tree. This approach could be used to detect species boundaries using genome divergence as a metric of the concordance between different loci in the genome. In cases of advanced speciation, a large proportion of the genome should show consistent phylogenetic patterns suggesting in turn sufficient time to accumulate private genetic differences (Schön *et al.*, 2009), purge ancestral polymorphism (Gao *et al.*, 2015; Guerrero & Hahn, 2018), and reduce the chance of gene flow (Dagilis *et al.*, 2022; Hamlin *et al.*, 2020). Mavengere *et al.* (2020) used 37 genomes in genome-level tests for genetic divergence (Mavengere *et al.*, 2020) confirming that the five phylogenetic species within *Paracoccidioides* fulfill all these requirements.

Previous assessments of genetic diversity have suggested that *Paracoccidioides* species have differences in their polymorphism level (Matute *et al.*, 2007; Mavengere *et al.*, 2020; Turissini *et al.*, 2017) but no study has addressed whether the demographic trajectories of *Paracoccidioides* species differ from each other. To this end, we used the pairwise sequentially Markovian coalescent (PSMC) method to compare the demographic patterns of two *Paracoccidioides* species: *P. restrepiensis* and *P. brasiliensis sensu stricto*. Our results suggest that, as previously proposed, *P. restrepiensis* has a lower population size as a consequence of a rapid decrease in its population size and genetic variability compared to *P. brasiliensis sensu stricto*. Here we speculate on the potential reasons behind this pattern of genetic variability.

## Methods

### Genomic data

To infer the changes in effective population size, we used the pairwise sequentially Markovian coalescent (PSMC) (Li & Durbin, 2011) method, which uses the genome of a single diploid individual per species. Since all species of *Paracoccidioides* are haploid, PSMC cannot be applied directly to the genome of a single isolate. Instead, we used two haploid individuals to generate a pseudodiploid genome. We generated two pseudodiploids for *P. restrepiensis*, one for *P. brasiliensis sensu stricto*, and one for *P. americana*. Although our focus was *P. restrepiensis*, the pseudodiploids from the two other species allowed us to compare it to the two other species. The Sequence Read Archive (SRA) numbers for the genomes used to generate pseudodiploids are listed in **table 1**.

**Table 1.** Genomes used to generate pseudodiploids for the PSMC analyses

Species	Genome 1	Genome 2
<i>P. restrepiensis</i>	EPM83	CAB
<i>P. restrepiensis</i>	H143	CNH
<i>P. americana</i>	T10B1	Pb02
<i>P. brasiliensis sensu stricto</i>	T1F1	PbBercelli

## Read mapping and variant calling

We mapped reads from the six resequenced genomes of the five known species to the *P. brasiliensis* strain Pb18 genome (BioProject accession number PRJNA28733 and BioSample accession number SAMN02953720) using the Burrows-Wheeler Aligner (BWA) version 0.7.12. This assembly currently has 57 supercontigs (Desjardins *et al.*, 2011). The resulting BAM files were merged using SAMtools version 0.1.19 for the variant call step. We remapped reads locally in the merged BAM files using the GATK version 3.2-2, RealignerTargetCreator, and IndelRealigner functions (DePristo *et al.*, 2011; McKenna *et al.*, 2010) and identified indels. Next, we identified single-nucleotide polymorphisms (SNPs) using the GATK UnifiedGenotyper function with the parameter “het” set to 0.01 and all others left as default. (The low *het* setting obeys to the fact that *Paracoccidioides* is haploid.) As in previous studies (Mavengere *et al.*, 2020), we filtered the Variant Call Format (VCF) file with the following parameters: QD=2.0, FS\_filter=60.0, MQ\_filter=30.0, MQ\_Rank\_Sum\_filter = -12.5, and Read\_Pos\_Rank\_Sum\_filter = -8.0. We excluded sites when their sequencing individual coverage was below the 5th quantile or above the 99th quantile of the genomic coverage distribution from any analyses.

### Phylogenetic tree

To infer divergence times between *P. restrepiensis* and its sister species *P. venezuelensis*, we used a genome-wide phylogenetic tree which inferred the genealogical relationships between the five known species of *Paracoccidioides* using a concatenated matrix of polymorphic sites. To convert tree distances to years, we used the mutation rate measured for *Saccharomyces* in mutation accumulation lines (Leffler *et al.*, 2012), an approach consistent with Turissini *et al.* (2017).

### Generation of pseudodiploids

PSMC uses levels of heterozygosity to infer population changes over time. Since *Paracoccidioides* is haploid, the method cannot be applied directly to assembled genomes. We used an alternative approach combining data from two individuals to generate a pseudo-diploid. To create pseudo-diploid sequences, we randomly selected heterozygous alleles from haploid genomes of two isolates from a sample of previously sequenced *Paracoccidioides* genomes (Mavengere *et al.*, 2020). We created pseudo-diploid sequences using the mergefa program in the seqtk package (<https://github.com/lh3/seqtk>) by merging haploid sequences of two individuals per species (Table 1). We only included sites with coverage between the 5th and 99th quantile of the genomic coverage distribution for both haploid isolates and if the SNP failed to pass one of the GATK filters.

### PSMC

PSMC calculates the effective population along slices of time using information from the rates of the coalescent events at a given time. To do this inference, we used the method developed by Li & Durbin (2011) (<https://github.com/lh3/psmc>). Briefly, the method uses the distribution of heterozygote sites across the genome and the pairwise sequentially Markovian coalescent (PSMC) method that defines a hidden Markov model where the parameters are the mutation rate, the recombination rate, and the effective population sizes through time. The parameters are inferred through an expectation-maximization algorithm. To assess the robustness of the inference, we did 100 bootstraps for each *Paracoccidioides* species. It should be noted that, unlike standard diploid PSMC plots, PSMC plots of pseudo-diploid genomes usually show an infinite effective population size at the point of divergence (Bazzicalupo *et al.*, 2022; Sato *et al.*, 2020). To convert generations to years, we used a substitution rate for nuclear DNA of  $1 \times 10^{-9}$  substitution/site/year following (Farlow *et al.*, 2015; Lynch *et al.*, 2008).

## Results and comments

### *Paracoccidioides restrepiensis* diverged from its sister species recently

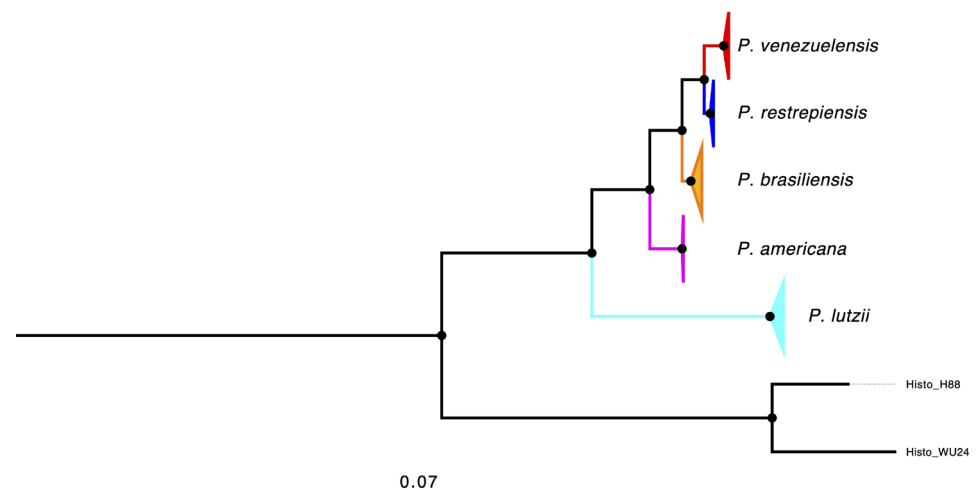
**Turissini et al.** (2017) used a multilocus sequence typing approach to infer the approximate divergence time of *Paracoccidioides* species. We expanded this analysis using the genome-wide phylogenetic tree inferred by **Mavengere et al.** (2020) (**Figure 1**). We found that the approximate age of divergence between *P. restrepiensis* and *P. venezuelensis* is 125,000 ( $\pm$  42,000) years, which is consistent with that inferred from a handful of nuclear loci and a constant substitution rate (**Turissini et al.**, 2017).

### *Paracoccidioides restrepiensis* has undergone a systematic population bottleneck

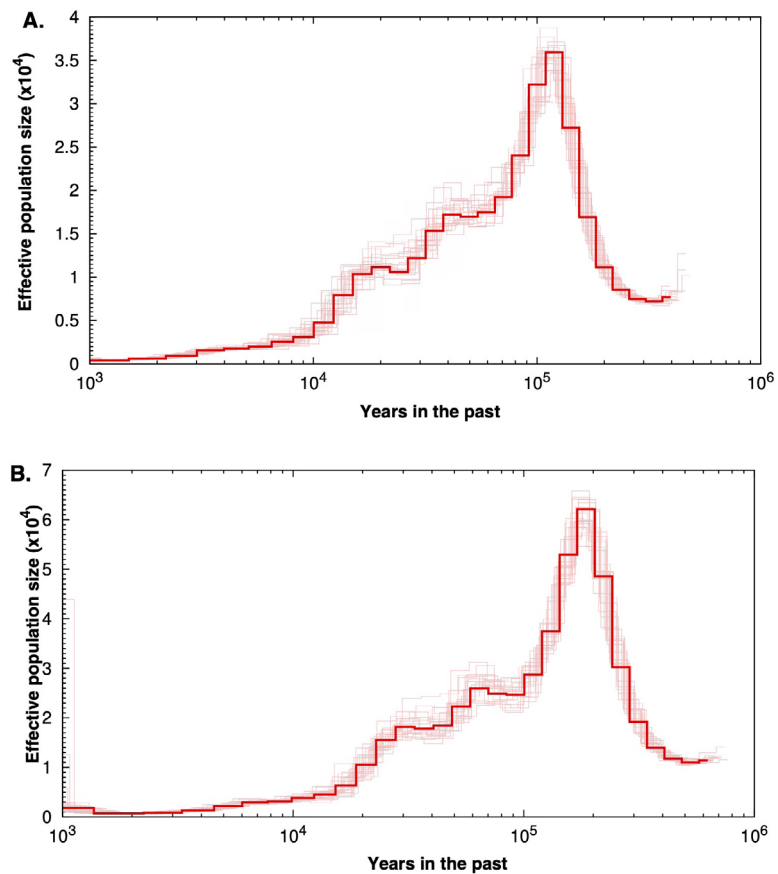
Previous analyses have shown that *P. restrepiensis* has a low level of genetic variation suggesting the possibility of a population bottleneck at some point in its evolutionary history. This pattern of low variability is consistent across the whole genome (**Mavengere et al.**, 2020). We investigated the trajectory of the effective population size in this species using PSMC in two different pseudodiploid genomes. The analyses showed a systematic reduction in the effective population size ( $N_e$ ) over the last 100,000 years (**Figure 2**); currently, the species shows its lowest historic  $N_e$ . This reduction in the  $N_e$  has a similar timeline to the divergence time from *P. venezuelensis* (**Turissini et al.**, 2017), *P. restrepiensis* sister species. Please note that although these dates are heavily affected by our assumptions of mutation rate and the number of generations per year, the pattern of constant  $N_e$  reduction remains unchanged.

### *Paracoccidioides brasiliensis sensu stricto* and *P. americana*

We used similar PSMC analyses with two other species of *Paracoccidioides* to assess if the pattern of  $N_e$  reduction over time was unique to *P. restrepiensis* or whether other species showed a similar trajectory. The analyses yielded two results that are noteworthy: First, we observed that for these other two species, the current effective population size is much larger than that of *P. restrepiensis*, which is in line with previous observations based on the level of heterozygosity (**Mavengere et al.**, 2020; **Muñoz et al.**, 2016; **Turissini et al.**, 2017). The higher level of these two species' genetic diversity is a historical trend and the two species have had a larger  $N_e$  than *P. restrepiensis* for at least the last million years. Second, neither of these two species shows the dramatic effective population size



**Figure 1.** Maximum likelihood tree depicting the genealogical relationships between *Paracoccidioides* species. *Paracoccidioides venezuelensis* is shown in red, *P. restrepiensis* in blue, *P. brasiliensis* in orange, *P. americana* in pink, and *P. lutzii* in cyan. All branches are supported by a bootstrap > 70%. Redrawn from **Mavengere et al.** (2020).



**Figure 2.** PSMC results for two pseudodiploid genomes of *P. restrepiensis*. The bolded line shows the median  $N_e$  (effective population size); other lines show 100 permutations. **A.** EPM83/CAB. **B.** H143/CNH. Note that both analyses show a similar trajectory.

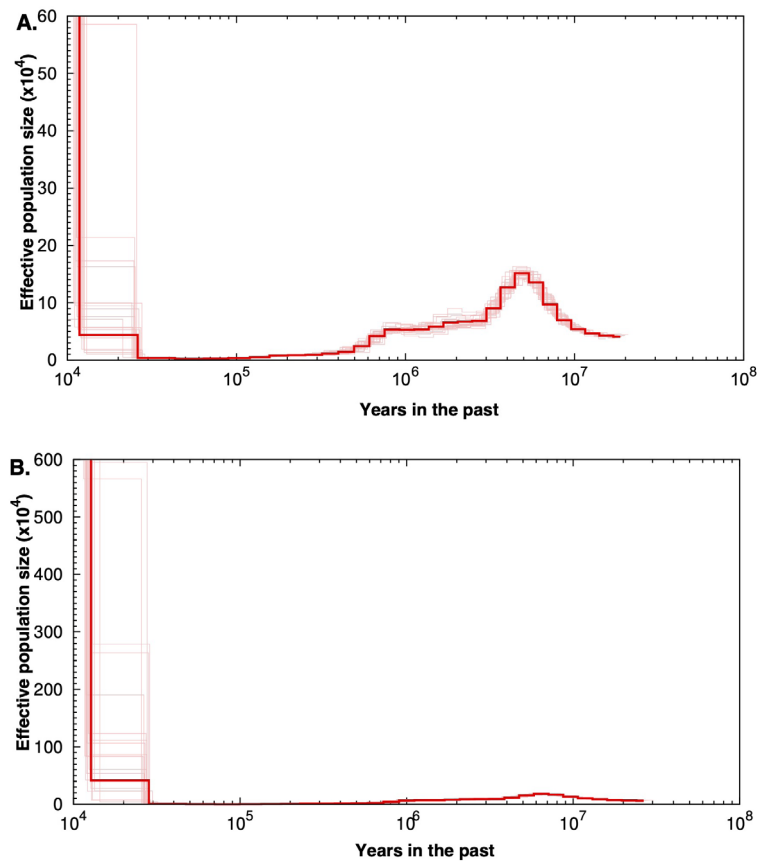
reduction observed in *P. restrepiensis*. The different trajectory of *P. restrepiensis* compared to that of the other *Paracoccidioides* species poses the question of the nature of biogeographic events that led to such a dramatic difference (**Figure 3**).

## Discussion

The evolutionary history of the different *Paracoccidioides* species remains a largely unstudied realm of the biology of the fungus. To date, most of the research has focused on describing the phylogenetic relationships between the different *Paracoccidioides* clades (**Matute et al., 2006; Teixeira et al., 2009; Teixeira et al., 2014**). In this report, we focused on the evolutionary dynamics within *Paracoccidioides* species and the trajectory of the  $N_e$  in *P. restrepiensis*. Our results suggest that *P. restrepiensis*, unlike other *Paracoccidioides* species, has undergone a continuous reduction of effective population size, which ultimately seems to have led to a genetically depauperate extant population.

Our results open two new research avenues regarding the biology of *P. restrepiensis*. First, recent collections suggested that *P. restrepiensis* is found not only in Colombia (**Cocio et al., 2020; Mattos et al., 2021; Teixeira et al., 2020**), which represents a formal test of the existence of isolation in the potential face of gene flow. More importantly, the expanded geographic range needs to be explored from the population genetics point of view. Multiple approaches have used the algorithm STRUCTURE to detect genetic partitions in the *Paracoccidioides* genus (**Bagagli et al., 2021**). These analyses have revealed the existence of genetic clusters consistent with the species delimitations from phylogenetic





**Figure 3.** PSMC results for two pseudodiploid genomes of *P. brasiliensis* (A) and *P. americana* (B). The bolded line shows the median  $N_e$ ; other lines show 100 permutations.

approaches. Nonetheless, they have not detected any population differentiation with *P. restrepiensis*. This absence of differentiation is not caused by the lack of power to detect structures within each of the phylogenetic species. For example, STRUCTURE detected the existence of two different clades of *P. brasiliensis sensu stricto*, S1a and S1b (Munoz *et al.*, 2016). Genome-wide tests for speciation revealed that the magnitude of genetic differentiation, albeit considerable, is not sufficient to classify these clades as independent species (Bagagli *et al.*, 2021; Mavengere *et al.*, 2020). The recent collection of *P. restrepiensis* in southern Brazil poses the question of whether this lack of structure and low historic effective population size are artifacts of focal collection in Colombia or whether the species is truly well mixed (Teixeira *et al.*, 2020).

One of the questions that arise concerns the sources of variation in *P. restrepiensis* because populations that have undergone extreme bottlenecks are more prone to extinction than those with large effective population sizes. Using full genome sequences and an array of statistical genetics, phylogenetics, and population genetics methods, previous studies have determined that *P. restrepiensis* evidences the exchange of a moderate number of alleles with other *Paracoccidioides* species similar to the levels observed among other *Paracoccidioides* species pairs. These observations are puzzling because the total amount of introgressed DNA was similar across the three species pairs and, given the different ages of the dyads and the expectations, there should be more hybrid incompatibilities in more diverged species pairs than in those recently diverged and, consequently, less possibility of introgression (Dagilis *et al.*, 2022). It is also puzzling that the amount of total introgression in these species pairs was comparable to that observed between species of the genus *Coccidioides* (*C. posadasii* and *C. immitis*), which diverged over 5

million years ago (Maxwell *et al.*, 2019). Whether *P. restrepiensis* can survive through a constant influx of introgression or through evolutionary rescue, “defined as the recovery of a population through natural selection”, is a question that needs to be addressed with a systematic approach.

Genetic divergence is an inexorable outcome of the passage of time. The recognition of species (using any of the available concepts) is not simply a scholastic exercise. On the contrary, the identification of species boundaries allows for the systematic exploration of clinical and epidemiological features that might distinguish between lineages. The use of distinct taxonomic names is also not set in stone. Names have fallen into disuse before (Lacaz *et al.*, 1997). The decision of giving a name to *P. restrepiensis* was not taken lightly; we studied the extent of differentiation at the genome level, at diagnostic loci (Matute *et al.*, 2006) and in morphological traits (Teixeira *et al.*, 2014; Turissini *et al.*, 2017). We also measured the extent of gene flow between species and found no evidence of introgression, a mark of an advanced level of speciation. The only stronger metric of speciation would be the measurement of reproductive isolation in controlled environmental conditions in a systematic crossing scheme. This experiment, however, is not possible for *Paracoccidioides*. The fungus is not known to reproduce sexually in laboratory conditions (even though the genome shows evidence of recombination in nature) (Matute *et al.*, 2006). Doing crosses between different species tends to be much more difficult than setting up crosses within species, a prospect that in *Paracoccidioides* is just staggering.

In the case of *P. restrepiensis*, there is no evidence that the fungus causes a disease that fundamentally differs from that caused by other *Paracoccidioides* species, although no study has sought such differences. These studies are cumbersome and will require a systematic assessment of a disease that is characterized by its long incubation period (Restrepo, 2000; Restrepo-Moreno *et al.*, 2020). Nonetheless, there are indications that *P. restrepiensis* differs from other species in clinically important traits. Serum IgG and IgE show different levels of reaction to antibody exposure in *P. lutzi*, *P. americana*, and *P. restrepiensis* (Assolini *et al.*, 2021). Similarly, even before the species boundaries were reported, *P. restrepiensis* was reported to have an unusually high level of antifungal resistance compared to isolates subsequently found to belong to the other *Paracoccidioides* species (Restrepo & Arango, 1980). The generality of these findings at the species level will require a full assessment of the phenotypic trait range across species.

Currently, there are few medically approved drugs for the treatment of mycoses; besides, the drugs have also been used for agricultural purposes for decades. The genetic information of pathogens is dynamic and they are constantly adapting to new hosts. Fungi such as *Paracoccidioides* spp. have highly efficient machinery (mutations, recombination, and horizontal gene transfer, among others) for genetic diversification and colonization of new environments (Taylor *et al.*, 2017). Emerging genomic variants associated with the antifungal resistance of other fungal species are being reported all over the world. Now, *P. restrepiensis* is a unique pathogenic fungal species circulating almost exclusively in Colombia and, therefore, it is a priority to continue with the genome surveillance and evolutionary history reconstruction of this endemic fungal species, particularly in a country with high biodiversity and active agricultural production as is the case of Colombia.

## Acknowledgments

This study received support from CODI-Universidad de Antioquia via a grant (*Sostenibilidad 2017/2018*) and a student scholarship from the post-graduation fund at Universidad de Antioquia.

## Author contribution

JCMCE: Writing of the original draft, review of the manuscript, writing of the final draft; OMG: Review of the draft and the manuscript; DRM: Writing of the main draft of the article, conceptualization and design of the methodology and data curation.



## Conflicts of interest

The authors declare no conflicts of interest.

## References

- Almeida, F. P.** (1930). Estudos comparativos do granuloma coccidióidico nos Estados Unidos e no Brasil. Novo gênero para o parasito brasileiro. *Annals Faculdade de Medicina Sao Paulo* 5, 125-141.
- Assolini, J. P., Lenhard-Vidal, A., Brecht, C. S. O., Tano, Z. N., Sano, A., Cezar-Dos-Santos, F., Ono, M. A., Itano, E. N.** (2021). Distinct Pattern of *Paracoccidioides lutzii*, *P. restrepiensis* and *P. americana* Antigens Recognized by IgE in Human Paracoccidioidomycosis. *Current Microbiology*, 78(7), 2608-2614. <https://doi.org/10.1007/s00284-021-02508-7>
- Bagagli, E., Matute, D. R., Garces, H. G., Tenorio, B. G., Garces, A. G., Alves, L. G. B., Yamauchi, D. H., Hrycyk, M. F., Barker, B. M., Teixeira, M. M.** (2021). *Paracoccidioides brasiliensis* Isolated from Nine-Banded Armadillos (*Dasypus novemcinctus*) Reveal Population Structure and Admixture in the Amazon Basin. *Journal of Fungi (Basel)*, 7(1). <https://doi.org/10.3390/jof7010054>
- Bazzicalupo, E., Lucena-Pérez, M., Kleinman-Ruiz, D., Pavlov, A., Trajçe, A., Hoxha, B., Sanaja, B., Gurielidze, Z., Kerdikoshvili, N., Mamuchadze, J., Yarovenko, Y. A., Akkiev, M. I., Ratkiewicz, M., Saveljev, A. P., Melovski, D., Gavashelishvili, A., Schmidt, K., Godoy, J. A.** (2022). History, demography and genetic status of Balkan and Caucasian *Lynx lynx* (Linnaeus, 1758) populations revealed by genome-wide variation. *Diversity and Distributions*, 28(1), 65-82. <https://doi.org/https://doi.org/10.1111/ddi.13439>
- Bellissimo-Rodrigues, F., Machado, A. A., Martinez, R.** (2011). Paracoccidioidomycosis epidemiological features of a 1,000-cases series from a hyperendemic area on the southeast of Brazil. *The American Journal of Tropical Medicine and Hygiene*, 85(3), 546-550. <https://doi.org/10.4269/ajtmh.2011.11-0084>
- Cocio, T.A., Nascimento, E., Kress, M., Bagagli, E., Martinez, R.** (2020). Characterization of a *Paracoccidioides* spp. strain from southeastern Brazil genotyped as *Paracoccidioides restrepiensis* (PS3) and review of this phylogenetic species. *Genetics and Molecular Biology*, 43(2), e20190201. <https://doi.org/10.1590/1678-4685-GMB-2019-0201>
- Coutinho, Z.F., Wanke, B., Travassos, C., Oliveira, R.M., Xavier, D.R., Coimbra, C. E., Jr.** (2015). Hospital morbidity due to paracoccidioidomycosis in Brazil (1998-2006). *Tropical Medicine and International Health*, 20(5), 673-680. <https://doi.org/10.1111/tmi.12472>
- Dagilis, A.J., Peede, D., Coughlan, J.M., Jofre, G.I., D'Agostino, E.R.R., Mavengere, H., Tate, A.D., Matute, D.R.** (2022). A need for standardized reporting of introgression: Insights from studies across eukaryotes. *Evolution Letters*, n/a(n/a). <https://doi.org/https://doi.org/10.1002/evl3.294>
- DePristo, M.A., Banks, E., Poplin, R., Garimella, K.V., Maguire, J.R., Hartl, C., Philippakis, A.A., del Angel, G., Rivas, M.A., Hanna, M., McKenna, A., Fennell, T.J., Kernysky, A.M., Sivachenko, A.Y., Cibulskis, K., Gabriel, S.B., Altshuler, D., Daly, M.J.** (2011). A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics*, 43(5), 491-498. <https://doi.org/10.1038/ng.806>
- Desjardins, C.A., Champion, M.D., Holder, J.W., Muszewska, A., Goldberg, J., Bailao, A.M., Brigido, M.M., Ferreira, M.E., Garcia, A.M., Grynberg, M., Gujja, S., Heiman, D.I., Henn, M.R., Kodira, C.D., Leon-Narvaez, H., Longo, L.V., Ma, L.J., Malavazi, I., Matsuo, A.L., . . . Cuomo, C. A.** (2011). Comparative genomic analysis of human fungal pathogens causing paracoccidioidomycosis. *PLoS Genetics*, 7(10), e1002345. <https://doi.org/10.1371/journal.pgen.1002345>
- Farlow, A., Long, H., Arnoux, S., Sung, W., Doak, T.G., Nordborg, M., Lynch, M.** (2015). The Spontaneous Mutation Rate in the Fission Yeast *Schizosaccharomyces pombe*. *Genetics*, 201(2), 737-744. <https://doi.org/10.1534/genetics.115.177329>
- Franco, M.F., Del Negro, G., Lacaz, C.d.S., Restrepo-Moreno, A.** (1994). *Paracoccidioidomycosis*. CRC Press.
- Gao, Z., Przeworski, M., Sella, G.** (2015). Footprints of ancient-balanced polymorphisms in genetic variation data from closely related species. *Evolution*, 69(2), 431-446. <https://doi.org/10.1111/evo.12567>
- Guerrero, R.F., Hahn, M.W.** (2018). Quantifying the risk of hemiplasy in phylogenetic inference. *Proceedings of the National Academy of Sciences USA*, 115(50), 12787-12792. <https://doi.org/10.1073/pnas.1811268115>

- Hamlin, J.A.P., Hibbins, M.S., Moyle, L.C.** (2020). Assessing biological factors affecting post-speciation introgression. *Evolution Letters*, 4(2), 137-154. <https://doi.org/10.1002/evl3.159>
- Lacaz Cda, S., Vidal, M.S., Pereira, C.N., Heins-Vaccari, E.M., de Melo, N.T., Sakai-Valente, N., Arriagada, G.L.** (1997). *Paracoccidioides cerebriformis* Moore, 1935. Mycologic and immunochemical study. *Rev Inst Med Trop Sao Paulo*, 39(3), 141-144. <https://doi.org/10.1590/s0036-46651997000300003>
- Leffler, E.M., Bullaughey, K., Matute, D.R., Meyer, W.K., Segurel, L., Venkat, A., Andolfatto, P., Przeworski, M.** (2012). Revisiting an old riddle: what determines genetic diversity levels within species? *PLoS Biology*, 10(9), e1001388. <https://doi.org/10.1371/journal.pbio.1001388>
- Li, H., Durbin, R.** (2011). Inference of human population history from individual whole-genome sequences. *Nature*, 475(7357), 493-496. <https://doi.org/10.1038/nature10231>
- Lutz, A.** (1908). Uma mycose pseudo-coccidica localizada na boca e observada no Brazil: contribuicao ao conhecimento das hyphoblastomycoses americanas. *Brasil Medico*, 22, 121-124, 141-144.
- Lutz, A.** (2004). *Lutz A Obra completa Uma micose pseudococcidica localizada na boca e observada no Brasil. Contribuição ao conhecimento das hifoblastomicoses americanas*. Fiocruz
- Lynch, M., Sung, W., Morris, K., Coffey, N., Landry, C.R., Dopman, E. B., Dickinson, W.J., Okamoto, K., Kulkarni, S., Hartl, D.L., Thomas, W.K.** (2008). A genome-wide view of the spectrum of spontaneous mutations in yeast. *Proceedings of the National Academy of Sciences USA*, 105(27), 9272-9277. <https://doi.org/10.1073/pnas.0803466105>
- Mattos, K., Cocio, T. A., Chaves, E. G. A., Borges, C.L., Venturini, J., de Carvalho, L.R., Mendes, R.P., Paniago, A.M.M., Weber, S.S.** (2021). An update on the occurrence of *Paracoccidioides* species in the Midwest region, Brazil: Molecular epidemiology, clinical aspects and serological profile of patients from Mato Grosso do Sul State. *PLoS Neglected Tropical Diseases*, 15(4), e0009317. <https://doi.org/10.1371/journal.pntd.0009317>
- Matute, D.R., McEwen, J.G., Puccia, R., Montes, B.A., San-Blas, G., Bagagli, E., Rauscher, J.T., Restrepo, A., Morais, F., Nino-Vega, G., Taylor, J.W.** (2006). Cryptic speciation and recombination in the fungus *Paracoccidioides brasiliensis* as revealed by gene genealogies. *Molecular Biology and Evolution*, 23(1), 65-73. <https://doi.org/10.1093/molbev/msj008>
- Matute, D.R., Sepulveda, V.E.** (2019). Fungal species boundaries in the genomics era. *Fungal Genetics and Biology*, 131, 103249. <https://doi.org/10.1016/j.fgb.2019.103249>
- Matute, D.R., Sepulveda, V.E., Quesada, L.M., Goldman, G.H., Taylor, J.W., Restrepo, A., McEwen, J.G.** (2006). Microsatellite analysis of three phylogenetic species of *Paracoccidioides brasiliensis*. *Journal of Clinical Microbiology*, 44(6), 2153-2157. <https://doi.org/10.1128/JCM.02540-05>
- Matute, D.R., Torres, I.P., Salgado-Salazar, C., Restrepo, A., McEwen, J.G.** (2007). Background selection at the chitin synthase II (chs2) locus in *Paracoccidioides brasiliensis* species complex. *Fungal Genetics and Biology*, 44(5), 357-367. <https://doi.org/10.1016/j.fgb.2007.01.006>
- Mavengere, H., Mattox, K., Teixeira, M.M., Sepulveda, V.E., Gomez, O.M., Hernandez, O., McEwen, J., Matute, D.R.** (2020). *Paracoccidioides* Genomes Reflect High Levels of Species Divergence and Little Interspecific Gene Flow. *mBio*, 11(6), e01999-01920. <https://doi.org/10.1128/mBio.01999-20>
- Maxwell, C.S., Mattox, K., Turissini, D.A., Teixeira, M.M., Barker, B.M., Matute, D.R.** (2019). Gene exchange between two divergent species of the fungal human pathogen, *Coccidioides*. *Evolution*, 73(1), 42-58. <https://doi.org/10.1111/evo.13643>
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., Garimella, K., Altshuler, D., Gabriel, S., Daly, M., DePristo, M.A.** (2010). The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Research*, 20(9), 1297-1303. <https://doi.org/10.1101/gr.107524.110>
- Morais, F.V., Barros, T.F., Fukada, M.K., Cisalpino, P.S., Puccia, R.** (2000). Polymorphism in the gene coding for the immunodominant antigen gp43 from the pathogenic fungus *paracoccidioides brasiliensis*. *Journal of Clinical Microbiology*, 38(11), 3960-3966. <http://jcm.asm.org/cgi/content/abstract/38/11/3960>
- Muñoz, J.F., Farrer, R.A., Desjardins, C.A., Gallo, J.E., Sykes, S., Sakthikumar, S., Misas, E., Whiston, E.A., Bagagli, E., Soares, C.M., Teixeira, M.M., Taylor, J. W., Clay, O.K., McEwen, J.G., Cuomo, C.A.** (2016). Genome Diversity, Recombination, and Virulence across the Major Lineages of *Paracoccidioides*. *mSphere*, 1(5), e00213-00216. <https://doi.org/10.1128/mSphere.00213-16>

- Niño-Vega, G.A., Calcagno, A.M., San-Blas, G., San-Blas, F., Gooday, G.W., & Gow, N.A. (2000). RFLP analysis reveals marked geographical isolation between strains of *Paracoccidioides brasiliensis*. *Medical Mycology*, 38(6), 437-441. <https://doi.org/10.1080/mmy.38.6.437.441>
- PAHO, (1971). Paracoccidioidomycosis. Proceedings of the First Pan American Symposium, Medellín, Colombia, Medellín, Colombia.
- Restrepo, A. (2000). Morphological aspects of *Paracoccidioides brasiliensis* in lymph nodes: implications for the prolonged latency of paracoccidioidomycosis?. *Medical Mycology*, 38(4), 317-322. <https://doi.org/10.1080/mmy.38.4.317.322>
- Restrepo, A., Arango, M. D. (1980). In vitro susceptibility testing of *Paracoccidioides brasiliensis* to sulfonamides. *Antimicrobial Agents and Chemotherapy*, 18(1), 190-194. <http://www.ncbi.nlm.nih.gov/pubmed/7416744>
- Restrepo-Moreno, A., Tobon-Orozco, A.M., González-Marín, A. (2020). Paracoccidioidomycosis. In J. E. Bennett, R. Dolin, & M. J. Blaser (Eds.), *Mandell, Douglas, and Bennett's principles and practice of infectious diseases* (Ninth edition. ed., Vol. 2, pp. 3211-3221). Elsevier.
- Sato, Y., Ogden, R., Kishida, T., Nakajima, N., Maeda, T., Inoue-Murayama, M. (2020). Population history of the golden eagle inferred from whole-genome sequencing of three of its subspecies. *Biological Journal of the Linnean Society*, 130(4), 826-838. <https://doi.org/10.1093/biolinnean/blaa068>
- Schön, I., Martens, K., Dijk, P.V. (2009). *Lost sex: the evolutionary biology of parthenogenesis*. Springer.
- Scorzoni, L., de Lucas, M.P., Singulani, J.L., de Oliveira, H.C., Assato, P.A., Fusco-Almeida, A.M., Mendes-Giannini, M.J.S. (2018). Evaluation of *Caenorhabditis elegans* as a host model for *Paracoccidioides brasiliensis* and *Paracoccidioides lutzii*. *Pathogens and Disease*, 76(1). <https://doi.org/10.1093/femspd/fty004>
- Siqueira, I.M., Fraga, C.L., Amaral, A.C., Souza, A.C., Jeronimo, M.S., Correa, J.R., Magalhaes, K.G., Inacio, C.A., Ribeiro, A.M., Burguel, P.H., Felipe, M.S., Tavares, A.H., Bocca, A.L. (2016). Distinct patterns of yeast cell morphology and host responses induced by representative strains of *Paracoccidioides brasiliensis* (Pb18) and *Paracoccidioides lutzii* (Pb01). *Medical Mycology*, 54(2), 177-188. <https://doi.org/10.1093/mmy/myv072>
- Soares, C., Madlun, E., da Silva, S., Pereira, M., Felipe, M. (1995). Characterization of *Paracoccidioides brasiliensis* isolates by random amplified polymorphic DNA analysis. *Journal of Clinical Microbiology*, 33(2), 505-507. <https://doi.org/10.1128/jcm.33.2.505-507.1995>
- Splendore, A. (1912). Zymonematosi con localizzazione nella cavita della bocca, osservata in Brasile. *Bulletin de la Société de Pathologie Exotique*, 5, 313-319.
- Taylor, J. W., Branco, S., Gao, C., Hann-Soden, C., Montoya, L., Sylvain, I., Gladieux, P. (2017). Sources of Fungal Genetic Variation and Associating It with Phenotypic Diversity. *Microbiology Spectrum*, 5(5). <https://doi.org/10.1128/microbiolspec.FUNK-0057-2016>
- Teixeira Mde, M., Theodoro, R.C., Oliveira, F.F., Machado, G.C., Hahn, R.C., Bagagli, E., San-Blas, G., Soares Felipe, M.S. (2014). *Paracoccidioides lutzii* sp. nov.: biological and clinical implications. *Medical Mycology*, 52(1), 19-28. <https://doi.org/10.3109/13693786.2013.794311>
- Teixeira, M.M., Cattana, M.E., Matute, D.R., Munoz, J.F., Arechavala, A., Isbell, K., Schipper, R., Santiso, G., Tracogna, F., Sosa, M. L. A., Cech, N., Alvarado, P., Barreto, L., Chacon, Y., Ortellado, J., Lima, C. M., Chang, M. R., Nino-Vega, G., Yasuda, M.A.S., ... Giusiano, G. (2020). Genomic diversity of the human pathogen *Paracoccidioides* across the South American continent. *Fungal Genetics and Biology*, 140, 103395. <https://doi.org/10.1016/j.fgb.2020.103395>
- Teixeira, M.M., Theodoro, R.C., de Carvalho, M. J., Fernandes, L., Paes, H.C., Hahn, R.C., Mendoza, L., Bagagli, E., San-Blas, G., Felipe, M.S. (2009). Phylogenetic analysis reveals a high level of speciation in the *Paracoccidioides* genus [Research Support, Non-U.S. Gov't]. *Mol Phylogenet Evol*, 52(2), 273-283. <https://doi.org/10.1016/j.ympev.2009.04.005>
- Teixeira, M.M., Theodoro, R.C., Nino-Vega, G., Bagagli, E., Felipe, M.S. (2014). Paracoccidioides species complex: ecology, phylogeny, sexual reproduction, and virulence. *PLoS Pathogens*, 10(10), e1004397. <https://doi.org/10.1371/journal.ppat.1004397>
- Turissini, D.A., Gómez, O.M., Teixeira, M.M., McEwen, J.G., Matute, D.R. (2017). Species boundaries in the human pathogen *Paracoccidioides*. *Fungal Genetics and Biology*, 106, 9-25. <https://doi.org/10.1016/j.fgb.2017.05.007>